

# **Short Note [Nota Corta]**

# PLANT PARASITIC NEMATODES ASSOCIATED WITH TREE TOMATO (Solanum betaceum Cav.) IN TLACOLULAN AND XALAPA, VERACRUZ, MEXICO†

[NEMATODOS FITOPARÁSITOS ASOCIADOS CON TOMATE DE ARBOL (Solanum betaceum Cav.) EN TLACOLULAN Y XALAPA, VERACRUZ, MÉXICO]

Gloria Carrión<sup>1</sup>, Lorena Velasco-Rodríguez<sup>2</sup> and Daniel López-Lima<sup>2\*</sup>

<sup>1</sup>Instituto de Ecología, A. C., Carretera Antigua a Coatepec 351, 91073 Xalapa, Veracruz, México. Email: <u>gloria.carrion@inecol.mx</u>

<sup>2</sup>Universidad Veracruzana, Facultad de Ciencias Agrícolas, Circuito Gonzalo Aguirre Beltrán s/n, 91000 Xalapa, Veracruz, México. Email: <u>danielopez@uv.mx</u>

\*Corresponding author

#### **SUMMARY**

Background: Tree tomato (Solanum betaceum Cav.) is a plant introduced to Mexico since colonial times, currently it is found as a crop in backyard orchards in the central mountainous region of Veracruz. Due to its nutraceutical properties, its cultivation is gaining importance worldwide and spreading. However, since there are no established formal crops in Mexico, there are no studies on the pests that may affect the development of this plant. Objective: To identify the plant-parasitic nematodes associated with the rhizospheric soil and root, as well as the determine their abundance in tree tomato plants, located in the Tlacolulan and Xalapa municipalities, in Veracruz state. Methodology: The nematodes were extracted from the soil and roots, fixed, clarified, mounted for identification at the genus level, and quantified. Results: 704 specimens belonging to 14 genera were identified: Criconema, Criconemoides, Filenchus, Fraglenchus, Gracilacus, Helicotylenchus, Malenchus, Meloidogyne, Ogma, Paratylenchus, Pratylenchus, Pratylenchoides, Sakia, and Thada. The most abundant nematodes were Helicotylenchus and Meloidogyne. Implications: Since in five samples we found Meloidogyne individuals, a diagnosis must be made prior to the establishment of new crops to avoid the increase in populations that can affect the performance of the plants. Conclusion: Criconema, Filenchus, Fraglenchus, Malenchus, Ogma, Pratylenchoides, Sakia, and Thada are recorded for the first time associated with this plant.

**Key words**: *Helicotylenchus*; *Meloidogyne*; Criconematidae; Tylenchidae; soil pests.

# RESUMEN

Antecedentes: El tomate de árbol (Solanum betaceum Cav.) es una planta introducida a México desde la época colonial, actualmente se encuentra como cultivo en huertas de traspatio en la región montañosa central de Veracruz. Debido a sus propiedades nutracéuticas, su cultivo está ganando importancia en el mundo y se está extendiendo. Sin embargo, dado que en México no existen cultivos formales establecidos, no hay información sobre las plagas que pueden afectar el desarrollo de esta planta. Objetivo: Identificar los nematodos fitoparásitos asociados al suelo rizosférico y raíz, así como determinar su abundancia en plantas de tomate de árbol, ubicadas en los municipios de Tlacolulan y Xalapa, en el estado de Veracruz. Metodología: Los nematodos fueron extraídos del suelo y raíces, fijados, aclarados, montados para su identificación a nivel de género y cuantificados. Resultados: Se identificaron 704 ejemplares pertenecientes a 14 géneros: Criconema, Criconemoides, Filenchus, Fraglenchus, Gracilacus, Helicotylenchus, Malenchus, Meloidogyne, Ogma, Paratylenchus, Pratylenchoides, Sakia y Thada. Los nematodos más abundantes fueron Helicotylenchus y Meloidogyne. Implicaciones: Dado que en cinco muestras encontramos individuos de Meloidogyne, se debe realizar un diagnóstico previo al establecimiento de nuevos cultivos para evitar el aumento de poblaciones que pueden afectar el rendimiento de las plantas. Conclusión: Criconema, Filenchus, Fraglenchus, Malenchus, Ogma, Pratylenchoides, Sakia y Thada se registran por primera vez asociados a esta planta.

Palabras clave: Helicotylenchus; Meloidogyne; Criconematidae; Tylenchidae; plagas del suelo.

Copyright © the authors. Work licensed under a CC-BY 4.0 License. https://creativecommons.org/licenses/by/4.0/

ORCID = G. Carrión https://orcid.org/0000-0003-0637-6108; D. López-Lima https://orcid.org/0000-0003-0523-8192

Submitted August 14, 2023 – Accepted October 16, 2023. <a href="http://doi.org/10.56369/tsaes.5095">http://doi.org/10.56369/tsaes.5095</a>

# INTRODUCTION

Solanum betaceum Cav. (Solanaceae) is native to southern Bolivia and northeastern Argentina, where it is found at altitudes between 1,100 and 2,300 meters above sea level (masl) (Bohs, 1994). During colonial times, the Spanish and Portuguese took it to various countries, and now, can be found plantations in various parts of the world (Lamas et al., 2022). Due to this distribution, it is known by various names such as mountain tomato, tamarillo, chilto, eggplant, and the main producing and exporting countries of this fruit are New Zealand, Portugal, Colombia, Ecuador, and Peru (Prohens et al., 1996; Ramírez and Kallarackal, 2019). This species has gained importance due to its nutraceutical properties, including the amount of polyphenols, flavonoids, carotenoids, anthocyanins, and vitamin C (Diep et al., 2020; Isla et al., 2022; Viera et al., 2022). Although it is not known when it was introduced to Mexico, there are records of specimens collected in the states of Chiapas (1945-1965), Guanajuato (1897), Jalisco (1886) and Veracruz (1976-1981) growing between 1,000 and 1,300 masl (Bohs, 1994). In the central region of Veracruz, the tree tomato has remained as a backyard crop and in the wild as part of the useful plants that grow under oak and cloud forests or in shaded coffee plantations. There are also small plantations, which have acquired commercial importance due to local and regional consumption and production prospects increasing (Villegas-Ruiz et al., 2013; Feicán-Mejía et al., 2016). In Ecuador and Colombia, where it is cultivated extensively and intensively, many aspects of tree tomato have been studied (Prohens and Nuez, 2001; Acosta-Quezada et al., 2011), including several pathogens (nematodes, fungi, bacteria, and viruses) that limit their production (Jaramillo et al., 2012; López-Cardona and Castaño-Zapata, 2013; Ramírez-Gil et al., 2017). In the Ecuador highlands, more than 10 genera of plant-parasitic nematodes associated with S. betaceum have been recorded, Meloidogyne, Nacobbus and Pratylenchus are the most frequent, causing in some cases losses greater than 90% and halving the useful life of the crop (Ramírez et al., 2015; Ubidia and Soria, 2017). Infected plants present symptoms such as nutrient deficiency, which is why on many occasions, the nematodes go unnoticed and proper management is not carried out (Wheeler et al., 2019). In Mexico, because S. betaceum is not intensively cultivated, there are no studies on its phytopathogens. However, considering the growing importance of this crop, it is necessary to study the possible pest organisms present in the areas where this plant grows, including plant-parasitic nematodes. In the present work, the genera of plant-parasitic nematodes associated with roots and rhizospheric soil of S. betaceum were identified and their abundance was determined.

#### MATERIALS AND METHODS

Soil and root samples were collected in October 2020 in the Tlacolulan and Xalapa municipalities, both in the Veracruz state, Mexico (Table 1). The sampled plants were found in backyard orchards or in the wild in the understory of the original vegetation (oak forest and cloud forest). At each sampling site, approximately 250 g of rhizospheric soil and 20 g of root tissue were taken. The samples were placed in plastic bags labeled with the location data of each plant and then taken to the laboratory. In total, 8 samples were taken in Tlacolulan and two in Xalapa.

For the extraction of ectoparasitic nematodes, 100 cm<sup>3</sup> of soil from each sample was processed using the sievingcentrifugation technique (van Bezooijen, 2006). For the extraction of endoparasitic nematodes, 10 g of root tissue from each sample was processed with the maceration, sieving, and centrifuging technique (van Bezooijen, 2006). The nematodes obtained from each sample were fixed with 4% boiling formalin and clarified by the Seinhorst method (1962). Subsequently, the specimens were placed in a Sedgwick-Rafter counting chamber and observed under a light microscope at 100X. The specimens with a stylet were mounted in a paraffin ring (van Bezooijen, 2006) on glass slides to observe them in the light microscope at 400 and 1000X and carry out the identification at the genus level based on morphological characters, such as stylet, esophagus, labial region, cuticle, reproductive organs and posterior region according to the taxonomic keys of the Tylenchida order (Siddiqi, 2000). Subsequently, the specimens were quantified, and the abundance of each genus was determined.

# RESULTS AND DISCUSSION

Plant parasitic nematodes obtained (704 specimens) from rhizospheric soil and tree tomato roots correspond to 14 genera: *Criconema, Criconemoides, Filenchus, Fraglenchus, Gracilacus, Helicotylenchus, Malenchus, Meloidogyne, Ogma, Paratylenchus, Pratylenchus, Pratylenchoides, Sakia,* and *Thada* (Table 1, Figure 1). From the total number of nematodes obtained, five were identified only at the family level (two Tylenchidae specimens) and subfamily (one Hoplolaiminae specimen and two Pratylenchinae specimens).

The nematodes associated with S. betaceum in this study coincide with those registered in the producing areas of Colombia and Ecuador where Criconemoides, Helicotylenchus. Meloidogyne, Paratylenchus Pratylenchus are present (Lozada et al., 2002; Ubidia & Soria, 2017). In addition, they have registered Heterodera. Nacobbus, Radopholus, Rotylenchus,

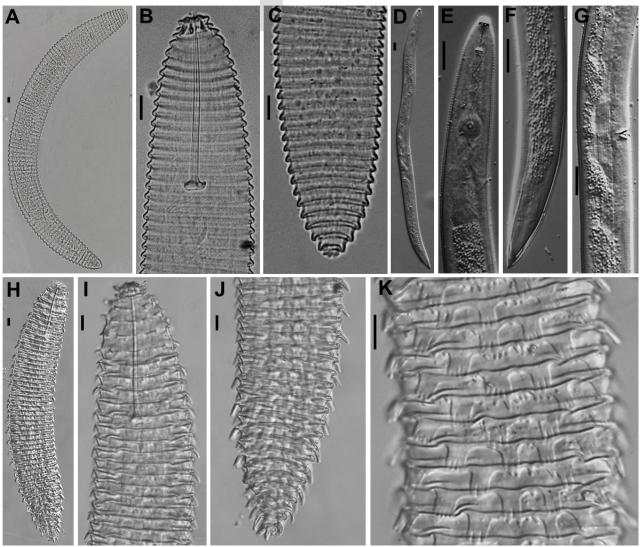
Table 1. Geographical data of collecting sites in tree tomato plants from Tlacolulan and Xalapa municipalities

(Veracruz, Mexico) and diagnosed genera.

Municipality	Sample	Location	Altitude masl	Cultivated/wild	Genera‡				
Tlacolulan	T1	19°39'51"N 97°00'07"W	1764	Backyard crop	Criconemoides, Filenchus, Gracilacus, Helicotylenchus, Malenchus, Meloidogyne, Pratylenchus				
	T2	19°39'53"N 97°00'21"W	1776	Wild	Helicotylenchus, Thada				
	Т3	19°40'3.69"N 97°00'12"W	1776	Backyard crop	Meloidogyne, Thada, Tylenchidae				
	T4	19°40'19"N 97°00'14"W	1840	Wild	Helicotylenchus, Fraglenchus, Malenchus, Pratylenchoides				
	T5	19°39'53"N 97°00'22"W	1777	Wild	Helicotylenchus				
	T6	19°40'02"N 97°00'12"W	1780	Backyard crop	Criconemoides, Helicotylenchus, Meloidogyne Ogma, Pratylenchus, Pratylenchinae				
	T7	19°39'53"N 97°00'15"W	1772	Wild	Helicotylenchus, Ogma				
	Т8	19°40'3.74"N 97°00'18"W	1846	Backyard crop	Criconema, Criconemoides, Helicotylenchus, Hoplolaiminae, Meloidogyne, Ogma, Paratylenchus, Tylenchidae				
Xalapa	X1	19°34'1.36"N 96°55'00"W	1402	Backyard crop	Criconemoides				
	X2	19°34'08"N 96°54'27"W	1381	Backyard crop	Malenchus, Meloidogyne, Sakia				

‡Based on morphological characters according to the taxonomic keys of the Tylenchida order (Siddiqi, 2000).

Tylenchus and Trichodorus (Ramírez et al., 2015; Mosquera-Espinosa, 2016). Likewise, in Kenya, Belonolaimus, Hemicycliophora, Longidorus, Paralongidorus, Paratrichodorus and Xiphinema have been recorded associated with this plant (Waswa et al., 2020). The difference in the genera registered in South America and those found in the present work may be due to the different environmental conditions in which the plants develop since in the areas of origin the studies were carried out on commercial crops and agricultural significantly influence the nematofauna (Guesmi-Mzoughi et al., 2022). On the other hand, in the present study the genera Criconema, Filenchus, Fraglenchus, Gracilacus, Malenchus, Ogma, Pratylenchoides, Sakia, and Thada are reported on S. betaceum for the first time. Criconema species parasitize 28 families of plants and have a preference for woody plants. It is suggested that crops susceptible to these nematode species should be grown away from forest areas (Rathore, 2019). It is likely that the presence of Criconema on S. betaceum is related to the presence of nearby trees. The species of the genus Filenchus are characterized by feeding on the epidermal cells of plant roots and fungal hyphae and may even feed on phytopathogenic fungi (Yeates et al., 1993; Okada et al., 2005; Munawar et al., 2022). However, there are no records of significant damage to crops, it is possible that they are found in the rhizosphere of the tree tomato feeding on fungi. The genus Gracilacus is common in undisturbed and cultivated soils, nonetheless, it is not considered of economic importance (Castillo et al., 1989). According to our bibliographical review, no significant damage to crops was reported. Malenchus species have a cosmopolitan distribution, their feeding habit is not well defined, but they are commonly found in association with decaying organic matter in the rhizosphere (Pedram et al., 2018), and no significant damage to crops was reported. Ogma species are commonly associated with trees such as *Pinus* spp. In particular, O. rhombosquamatum causes thickening of root cortical cells, leading to necrosis in the olive trees epidermis, all the same, it is not considered of economic importance (Vovlas and Inserra, 1981). Pratylenchoides species are migratory endoparasites on several crops (Ghaderi and Karegar, 2014). These nematodes have been found to affect the roots of Solanum lycopersicum and Perilla frutescens (Li et al., 2020; Kang et al., 2023). Tomato trees may be susceptible to Pratylenchoides; yet, pathogenicity studies are needed to determine its potential as a pest. Regarding the genera Fraglenchus, Sakia and Thada, they have been found in the rhizosphere of several plants, but there is no definitive information about their feeding habits (Siddiqi, 2000) and, according to our bibliographic review, they are not considered agricultural pests.



**Figure 1.** Criconemoides sp. female: A) entire body, B) body anterior region, C) posterior region. Pratylenchoides sp. female: D) entire body, E) body anterior region, F) posterior region, G) lateral field with six incisures. Ogma sp. female: H) entire body, I) anterior region, J) posterior region, K) body annules with finger-like projections (Scale bars =  $10 \mu m$ ). All photographs of the nematodes are original and were taken by the authors.

The highest abundance in rhizospheric soil was found in sample 5 from Tlacolulan, with 200 specimens per 100 cm<sup>3</sup> soil<sup>-1</sup>, all the genus *Helicotylenchus*, followed by sample 6 from the same locality with 97 specimens per 100 cm<sup>3</sup> soil<sup>-1</sup> of which 80 were *Helicotylenchus*, 16 *Criconemoides* and one *Meloidogyne* specimen. Likewise, sample 1 of Tlacolulan presented the highest *Meloidogyne* abundance in soil, with 30 specimens, followed by *Criconemoides* and *Filenchus* with 24 and 14 specimens respectively (Table 2). The highest abundance in the roots was found in sample 2 from Xalapa with 116 specimens per 10 g of which 113 correspond to *Meloidogyne* and 3 to *Malenchus*. Likewise, samples 5 and 6 of Tlacolulan presented a

density of 31 and 46 nematodes per 10 g root<sup>-1</sup> where the most abundant genus was *Helicotylenchus*, coinciding with the soil samples from the same sites (Table 3).

The most abundant genus in this work was *Helicotylenchus*, which contrasts with the tree tomatoproducing areas of Colombia (Mosquera-Espinosa, 2016), Ecuador (Ubidia and Soria, 2017), Kenya (Juma *et al.*, 2020; Waswa *et al.*, 2020), and New Zealand (Knight, 2001) where the highest abundance corresponds to the *Meloidogyne* genus. This could be due to the monoculture effect, which provides favorable conditions for *Meloidogyne* reproduction such as low biological diversity in the soil. Likewise, some of the most affected

plantations were established in fields where vegetables, coffee, plantain, lulo and other plants highly susceptible to *Meloidogyne* were previously grown (Lozada *et al.*, 2002; García *et al.*, 2004; Mosquera-Espinosa, 2016). In contrast, the plants studied in the present work were found in backyard orchards or growing wild, which allows the regulation of plant-parasitic nematode abundance, due to the greater biological diversity in the soil (Topalovic *et al.*, 2020).

Although *Meloidogyne* J2 juveniles were found in five of the samples studied, no significant root damage was observed, even in sample 2 from Xalapa where a high infestation level was recorded. It is necessary to carry out the *Meloidogyne* species determination in the plants and their pathogenicity. On the other hand, to date, there are no studies to calculate the economic threshold regarding the nematode infestation in *S. betaceum* (Lozada *et al.*, 2002). In general, for *Meloidogyne*, between 1 and 5 individuals per gram of soil are considered to cause losses in various Solanaceae species (Firoza and Maqbool, 1995; Evlice *et al.*, 2021; Talavera-Rubia *et al.*, 2022). According to these data, all the samples where *Meloidogyne* individuals were detected could be affected. However, it is necessary to carry out studies to determine the tolerance to the different plant-parasitic nematodes genera in *S. betaceum* and to establish an economic threshold for this plant.

Table 2. Plant parasitic nematode abundance in 100 cm<sup>3</sup> of *Solanum betaceum* rhizospheric soil in the Tlacolulan (T)

and Xalapa (X) municipalities, Veracruz, Mexico.

Family/Genera	Site									
-	<b>T1</b>	<b>T2</b>	<b>T3</b>	<b>T4</b>	T5	<b>T6</b>	<b>T7</b>	<b>T8</b>	<b>X1</b>	<b>X2</b>
Criconemoides	24					16		7	1	
Filenchus	14									
Gracilacus	1									
Helicotylenchus	3	2		52	200	80	6	3		
Malenchus										1
Meloidogyne	30					1		6		
Paratylenchus								5		
Sakia										1
Thada		2	1							
Tylenchidae			1							
Total	71	4	2	52	200	97	6	21	1	2

Table 3. Plant parasitic nematode abundance in 10 g root-1 of *Solanum betaceum* in the Tlacolulan (T) and Xalapa

(X) municipalities, Veracruz, Mexico.

Family/Genera	Site									
	<b>T1</b>	<b>T2</b>	<b>T3</b>	<b>T4</b>	<b>T5</b>	<b>T6</b>	<b>T7</b>	T8	<b>X1</b>	<b>X2</b>
Criconema	_							1		
Criconemoides						1		2		
Fraglenchus				2						
Helicotylenchus	1				31	40		8		
Hoplolaiminae								1		
Malenchus	1			6						3
Meloidogyne			2					1		113
Ogma						2	1	1		
Paratylenchus								15		
Pratylenchus	1					1				
Pratylenchinae						2				
Pratylenchoides				11						
Tylenchidae								1		
Total	3	0	2	19	31	46	1	30	0	116

#### **CONCLUSIONS**

The diagnosis of plant-parasitic nematodes is difficult because it is not possible to perform visual evaluations in the aerial parts of the plants since the damage is caused in the roots. Making a proper diagnosis before establishing new crops or undertaking management actions is important. In the municipalities of Tlacolulan and Xalapa, Veracruz, Mexico, a diversity of plant-parasitic nematodes was found associated with the rhizospheric soil and the roots of S. betaceum. Criconema, Filenchus, Fraglenchus, Malenchus, Ogma, Pratylenchoides, Sakia, and Thada are recorded for the first time associated with S. betaceum. In most of the samples, the nematode abundance was low to affect plant development. However, we recommend carrying out a diagnosis prior to the establishment of crops to prevent the increase in populations that could affect plant yield.

**Funding.** No financial support was received to carry out this work.

**Conflict of interest.** The authors declare that they have no competing interests.

Compliance with ethical standards. The authors confirm that this investigation was conducted under the current ethical procedures. No humans or animals were used in the studies of this article.

**Data availability.** Data is available with the corresponding author upon request.

Author contribution statement (CRediT). G. Carrión: Conceptualization, Writing, Methodology, Supervision. L. Velasco-Rodríguez: Writing, Sample acquisition, Methodology. D. López-Lima: Conceptualization, Investigation, Writing, Validation.

# REFERENCES

- Acosta-Quezada, P.G., Martínez-Laborde, J.B. and Prohens, J., 2011. Variation among tree tomato (*Solanum betaceum* Cav.) accessions from different cultivar groups: implications for conservation of genetic resources and breeding. *Genetic Resources and Crop Evolution*, 58(6), pp. 943–960. <a href="https://doi.org/10.1007/s10722-010-9634-9">https://doi.org/10.1007/s10722-010-9634-9</a>
- Bohs, L., 1994. Cyphomandra (Solanaceae). *Flora Neotropica*, 63, pp. 1–175. http://www.jstor.org/stable/4393848
- Castillo, P., Gonzalez-Pais, M.A. and Gomez-Barcina, A., 1989. El género *Gracilacus* Raski, 1962 en España (Paratylenchidae: Tylenchida). *Revista*

- *Ibérica de Parasitologia*, 49(4), pp. 321-328. <a href="https://www.plant-animal.es/pdfscazorla/CH48.pdf">https://www.plant-animal.es/pdfscazorla/CH48.pdf</a>
- Diep, T.T., Pook, C., Rush, E. and Yoo, M.J., 2020. Quantification of Carotenoids, α-Tocopherol, and Ascorbic Acid in Amber, Mulligan, and Laird's Large Cultivars of New Zealand Tamarillos (*Solanum betaceum* Cav.). *Foods*, 9(6), p. 769. https://doi.org/10.3390/foods9060769
- Evlice, E., Toktay, H., Yatkın, G., Erdoğuş, F.D. and Imren, M., 2021. Population fluctuations of root-knot nematodes *Meloidogyne chitwoodi* and *M. hapla* under field conditions. *Phytoparasitica* 50, pp. 233–242. https://doi.org/10.1007/s12600-021-00939-3
- Feicán-Mejía, C.G., Encalada-Alvarado, C.R. and Becerril-Román, A.E., 2016. Descripción agronómica del cultivo de tomate de árbol (*Solanum betaceum* Cav.). *Agroproductividad*, 9(8), pp. 78-86. <a href="https://revista-agroproductividad.org/index.php/agroproductividad/article/view/806">https://revista-agroproductividad.org/index.php/agroproductividad/article/view/806</a>
- Firoza, K. and Maqbool, M.A., 1995. Numerical threshold for infection of the spiral nematode, *Helicotylenchus dihystera* (Cobb, 1893) Sher, 1961 on brinjal, tomato and wheat. *Pakistan Journal of Nematology*, 13(2), pp. 93-97. <a href="https://www.pjn.com.pk/pjn/files/vol%2013%20">https://www.pjn.com.pk/pjn/files/vol%2013%20</a> no.%202/6.pdf
- García, F., Obando, J. and Betancourt, G.C., 2004.

  Reconocimiento de especies de *Meloidogyne* en tomate de árbol (*Solanum betacea*) y lulo (*Solanum quitoense*) en la zona norte del Departamento de Nariño. *Revista de Ciencias Agrícolas*, 21(1), pp. 28-40.

  <a href="https://dialnet.unirioja.es/servlet/articulo?codigo=6191592">https://dialnet.unirioja.es/servlet/articulo?codigo=6191592</a>
- Ghaderi, R. and Karegar, A., 2014. Contribution to a revision of the genus *Pratylenchoides* Winslow, 1958 (Nematoda: Merliniidae), with redescription of *P. erzurumensis* Yüksel, 1977 from Iran. *Zootaxa* 3900(3), pp. 339-369. https://doi.org/10.11646/zootaxa.3900.3.2
- Guesmi-Mzoughi, I., Tabib, M., Sellami, F., Hadj-Naser, F., Regaieg, H., Kallel, S. and Horrigue-Raouani, N., 2022. Diversity of plant-parasitic nematode communities infesting olive orchards in Tunisia in relation to agronomic factors. *European Journal of Plant*

- Pathology, 164(4), pp. 479–494. https://doi.org/10.1007/s10658-022-02572-0
- Isla, M.I., Orqueda, M.E., Moreno, M.A., Torres, S. and Zampini, I.C., 2022. *Solanum betaceum* fruits waste: a valuable source of bioactive compounds to be used in foods and non-foods applications. *Foods*, 11(21), pp. 3363. https://doi.org/10.3390/foods11213363
- Jaramillo, Z.M.M., Álvarez, J.A. and Marín, M.M., 2012.
  Características de los virus asociados a la virosis del tomate de árbol (*Solanum betaceum*) en Colombia. *Revista Lasallista de Investigación*, 9(1), pp. 115-127.
  <a href="http://www.scielo.org.co/scielo.php?script=sci">http://www.scielo.org.co/scielo.php?script=sci</a> a rttext&pid=S1794-44492012000100012
- Juma, W.S., Waceke, J.W., and Nchore, S.B., 2020. Diversity of plant parasitic nematodes of tree tomato (*Solanum betaceum* Cav.) in Kiambu and Embu Counties, Kenya. *Middle East Journal*, 9(3), pp. 605-616. http://doi.org/10.36632/mejar/2020.9.3.47
- Kang, H., Ko, H-R., Park, B-Y., Park, S., Park, E. and Kim, E., 2023. First report of *Pratylenchoides leiocauda* parasitic on *Perilla frutescenes* in the Republic of Korea. *Plant Disease*, 107(1), pp. 234. <a href="https://doi.org/10.1094/PDIS-10-21-2322-PDN">https://doi.org/10.1094/PDIS-10-21-2322-PDN</a>
- Knight, K.W., 2001. Plant parasitic nematodes associated with six subtropical crops in New Zealand. *New Zealand Journal of crop and horticultural science*, 29(29), pp. 267-275. <a href="https://doi.org/10.1080/01140671.2001.9514188">https://doi.org/10.1080/01140671.2001.9514188</a>
- Lamas, C.Y., Urtasun, M.M., Giamminola, E.M., Pratta, G.R., Caruso, G., Morandini, M., and De Viana, M.L., 2022. Fruit and seed characterization of wild populations of a traditional Andean crop: *Solanum betaceum* Cav. (Solanaceae) in the Argentinian Yungas. *Genetic Resources and Crop Evolution*, 69(1), pp. 231–244. https://doi.org/10.1007/s10722-021-01223-3
- Li, R., Gu, J.F., Li, H.M., Zhang, X.M. and Wang, X., 2020. Morphological and molecular identification of *Pratylenchoides leiocauda* in tomato. *Journal of Nanjing Agricultural University*, 43(6), pp. 1033-1041. https://doi.org/10.7685/jnau.202003011

- López-Cardona, N. and Castaño-Zapata, J., 2013. Etiología de la muerte descendente del tomate de árbol [Solanum betaceum (Cav.) sendt.]. Revista Agronomía, 21(1), pp. 7-18. https://www.doc-developpement-durable.org/file/Culture/Arbres-Fruitiers/FICHES ARBRES/Tamarillo tomate arbustive/ETIOLOG%C3%8DA%20DE%20LA%20MUERTE%20DESCENDENTE%20DEL%20TOMATE%20DE%20%C3%81RBOL.pdf
- Lozada, S.L., Varón de A, F. and Gómez, E.D., 2002. Nematodos asociados al tomate de árbol *Solanum betaceum* en el Valle del Cauca. *Fitopatología Colombiana*, 26(2), pp. 126-134. https://agriperfiles.agri-d.net/display/n44101
- Mosquera-Espinosa, A.T., 2016. Fitonematodos asociados a *Cyphomandra betacea* (Cav.) Sendtn., *Solanum quitoense* Lam. y *Daucus carota* L. en el Departamento de Boyacá, Colombia. *Acta Agronómica*, 65(1), pp. 87–97. <a href="http://dx.doi.org/10.15446/acag.v65n1.45180">http://dx.doi.org/10.15446/acag.v65n1.45180</a>
- Munawar, M., Castillo, P. and Yevtushenko, D.P., 2022.

  Description of *Filenchus* Species from Agroecosystem of Southern Alberta, Canada. *Agronomy*, 12(3), pp. 690. https://doi.org/10.3390/agronomy12030690
- Okada, H., Harada, H. and Kadota, I., 2005. Fungal-feeding habits of six nematode isolates in the genus *Filenchus. Soil Biology & Biochemistry*, 37(6), pp. 1113–1120. https://doi.org/10.1016/j.soilbio.2004.11.0
- Pedram, M., Soleymanzadeh, M., Pourjam, E. and Mobasserim, M., 2018. Observations on *Malenchus geraerti* n. sp. (Rhabditida: Tylenchidae), a morphological and molecular phylogenetic study. *Zootaxa* 4369(3), pp. 406-418. https://doi.org/10.11646/zootaxa.4369.3.6
- Prohens, J. and Nuez, F., 2001. The Tamarillo (*Cyphomandra betacea*), A review of a promising small fruit crop. *Small Fruits Review*, 1(2), pp. 43-68. http://doi.og/10.1300/j301v01n02\_06
- Prohens, J., Ruiz, J. J. and Nuez, F., 1996. Advancing the tamarillo harvest by induced postharvest ripening. *HortScience*, 32(1), pp. 109-111. https://doi.org/10.21273/HORTSCI.31.1.109
- Ramírez-Gil, J.G., Gil-Aguirre, A. and Morales-Osorio, J.G., 2017. Etiology of tree tomato (*Solanum*

- betaceum Cav.) diseases. Revista de Protección Vegetal, 32(1), pp. 33-51. http://scielo.sld.cu/pdf/rpv/v32n1/rpv04117.pdf
- Ramírez, F. and Kallarackal, J., 2019. Tree tomato (Solanum betaceum Cav.) reproductive physiology: A review. Scientia horticulturae, 248, pp. 206-215. https://doi.org/10.1016/j.scienta.2019.01.019
- Ramírez, F., Grijalva, R., Navarrete, X. and Guerrero, R., 2015. Nematodos fitoparásitos asociados con tomate de árbol (*Solanum betaceum* Cav.) en las provincias de Imbabura, Pichincha y Tungurahua, Ecuador. *Ecuador Es Calidad*, 2(1), pp. 1-6. https://doi.org/10.36331/revista.v2i1.2
- Rathore, Y.S., 2019. A note on host diversity ok *Criconema* spp. *Pantnagar Journal of Research*, 17(3), pp. 280-282. <a href="https://www.gbpuat.res.in/uploads/archive/17.3.17.pdf">https://www.gbpuat.res.in/uploads/archive/17.3.17.pdf</a>
- Seinhorst, J., 1962. On the Killing, Fixation and Transferring to Glycerin of Nematodes. *Nematologica*, 8(1), pp. 29-32. https://doi.org/10.1163/187529262x00981
- Siddiqi, M.R., 2000. *Tylenchida: Parasites of Plants and Insects*. Wallinford: CAB International.
- Talavera-Rubia, M., Vela-Delgado, M.D. and Verdejo-Lucas, S., 2022. A cost-benefit analysis of soil disinfestation methods against root-knot nematodes in mediterranean intensive horticulture. *Plants*, 11, pp. 2774. https://doi.org/10.3390/plants11202774
- Topalović, O., Hussain, M. and Heuer, H., 2020. Plants and Associated Soil Microbiota Cooperatively Suppress Plant-Parasitic Nematodes. *Frontiers in Microbiology*, 11, pp. 313. https://doi.org/10.3389/fmicb.2020.00313
- Ubidia, V.P. and Soria, C., 2017. Nematodos parásitos asociados con tomate de árbol (*Solanum betaceum* Cav.) en la sierra del Ecuador. *Revista Ecuatoriana De Medicina Y Ciencias Biológicas* (*REMCB*), 38(2), pp. 107-118. https://doi.org/10.26807/remcb.v38i2.549

- van Bezooijen, J., 2006. *Methods and techniques for nematology*. Wageningen University. <a href="https://www.wageningenur.nl/en/show/Manual-Methods-and-Techniques-for-nematology-1.htm">https://www.wageningenur.nl/en/show/Manual-Methods-and-Techniques-for-nematology-1.htm</a>
- Viera, W., Samaniego, I., Camacho, D.M.A., Habibi, N., Ron, L., Sediqui, N., Álvarez, J.S., Viteri, P., Sotomayor, A., Merino, J.V., Vásquez-Castillo, W. and Brito, B., 2022. Phytochemical characterization of a tree tomato (*Solanum betaceum* Cav.) breeding population grown in the Inter-Andean Valley of Ecuador. *Plants*, 11(3), pp. 268. https://doi.org/10.3390/plants11030268
- Villegas-Ruíz, X., Rodríguez-Armas, D.N., Guerrero-Beltrán, J.Á. and Bárcenas-Pozos, M.E., 2013. Estabilidad de un producto dulce de tamarillo (*Cyphomandra betacea*) conservado por métodos combinados. *Scientia Agropecuaria*, 4(2), pp. 89-100. https://www.redalyc.org/articulo.oa?id=357633705002
- Vovlas, N. and Inserra, R.N., 1981. Parasitic habits of Ogma rhombosquamatum and description of the male. Journal of Nematology, 13(1), pp. 87-90. <a href="https://www.ncbi.nlm.nih.gov/pmc/articles/PMC">https://www.ncbi.nlm.nih.gov/pmc/articles/PMC</a> 2618052/pdf/87.pdf
- Waswa, S.J., Waceke, J.W. and Nchore, S.B., 2020. Diversity of plant parasitic nematodes of tree tomato (*Solanum betaceum* Cav.) in Kiambu and Embu Counties, Kenia. *Middle East Journal of Agriculture Research* 9(3), pp. 605-616. https://doi.org/10.36632/mejar/2020.9.3.47
- Wheeler, D.A., Scott, J.C., Dung, J.K.S. and Johnson, D.C., 2019. Evidence of a trans-kingdom plant disease complex between a fungus and plant-parasitic nematodes. *PLOS ONE*, 14(2), pp. e0211508. <a href="https://doi.org/10.1371/journal.pone.0">https://doi.org/10.1371/journal.pone.0</a> 211508
- Yeates, G.W., Bongers, T., De Goede, R.G., Freckman, D.W. and Georgieva, S.S., 1993. Feeding habits in soil nematode families and genera—An outline for soil ecologists. *Journal of Nematology*, 25(3), pp. 315–331. <a href="https://www.ncbi.nlm.nih.gov/pmc/articles/PMC">https://www.ncbi.nlm.nih.gov/pmc/articles/PMC</a> 2619405/pdf/315.pdf